Production of 3rd generation biofuel from green microalgae : *Oedogonium nodulosum*

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Abstract— The socio-economic and environmental situation strongly marked by the increase in oil production and global warming has led researchers towards an alternative path such as renewable energies. The purpose of this study was to synthesize biofuel from oil extracted from green microalgae by the transesterification process using ethanol as alcohol and potassium hydroxide (KOH) as homogeneous catalyst for the reaction. Odogonium nodulosum was the selected green microalgae for our Biodiesel production. This species exhibited high capacity of lipid (45.38% in dry weight) which could be of high potential as raw materials for biofuel production, with an overall efficiency of 80%. Some physic-chemical properties and characterization of the obtained products were also verified such relative density, refractive index, kinematic viscosity, cloud point, color and Biodiesel flash point measurements. The comparison of our results with the properties of Diesel and Gasoline showed us that our biodiesel has properties very similar to those of diesel but more ecological.

Keywords— Green microalgae, *Odogonium nodulosum*, Lipids extraction, Transesterification, Biofuel

I. INTRODUCTION

With the continued development of industry and transport, energy demand in our world is increasing day after day. Oil, or even all fossil fuels (coal, natural gas and oil), currently provide almost all of this energy. However, these resources are not renewable and their consumption at the current rate will lead to the disappearance of reserves in a few decades [1]. On a global scale, the solar energy received represents about 10,000 times humanity's annual energy consumption. It is clearly the most abundant source of renewable energy [2]. Photosynthesis is an excellent natural way to convert and store solar energy [3]. The world of aquatic photosynthetic microorganisms, including microalgae, represents considerable and almost untapped potential for bioenergy production [4, 5]. Indeed, these microorganisms are of great importance, due to their ability to produce high-value and energy-interesting compounds such as: fats, hydrogen, carbohydrates and proteins [6, 7]. Their use makes it possible to consider the development of innovative biofuel production processes [8].

Biofuels are intended to replace petroleum-based fuel. They are a sustainable alternative to fossil fuels as they are renewable and less toxic to the environment. For the time being, these biofuels are of vegetables origin and come from agricultural crops. A major disadvantage is the debate between the use of land to produce food or biofuel [9]. As a result, many studies have been conducted to use an alternative energy source, many of which focus on algae.

Indeed, the biomass from algae is convertible via chemical or enzymatic transformations into biodiesel or bioethanol. Almost all biomass can be transformed into transport energy. The lipids are converted into biodiesel and the rest of the biomass can be used to produce bio-methanol. The production of biofuel from algae has many advantages that go beyond replacing oil as transport energy [10, 11].

In fact, the purpose of this study is to synthesize a biodiesel from green microalgae oils by transestérification using a basic homogenous catalyst.

The most effective basic catalysts used for the transesterification of triglycerides include NaOH, NaOMe, KOH and KOMe [12, 13]. In addition, the basic catalyst technique in a homogeneous medium in the presence of methanol (MeOH), is the most commonly used because it is the cheapest and can be carried out at low temperatures and pressures [14].

II. METHODS AND MATERIALS

Microalgae samples were collected from a freshwater channel covered with filamentous algae according to the CIPEL protocol [15]. The samples were grown at laboratory scale in an algal photobioreactor in batch mode. A dominant green algae species has been selected for the production of biofuel. It is *Oedogonium nodulosum* (Fig. 1). The most important physical (light, agitation, temperature) and chemical (various nutrients) factors necessary for its growth and the culture medium used were considered [16]. This microalgae was initially grown in batch mode in 1.5 liter photobioreactors under sterile and aseptic conditions, while considering that light and carbon dioxide have an important influence on the cell growth of *Oedogonium nodulosum*.



Fig. 1 Oedogonium nodulosum (x400 magnification)

A biomass-concentration monitoring was carried out in order to recover the optimal amount of our selected species under optimal growth conditions (temperature (22°C), light (120 μ mol.m⁻².s⁻¹), agitation (100 rpm), dissolved CO₂ level (13%), pH \approx 6 to 8, and nutrients). The maximum biomass growth rate μ max is calculated as follows:

$$\mu_{max} = \frac{\log_2 N_n - \log_2 N_0}{t_n - t_0}$$

With :

 μ_{max} : maximum biomass growth rate (mn^{-1}) ; log_2 : decimal logarithm; N_0 : Initial concentration of dry algal mass $(mg.L^{-1})$ that corresponds to the initial time t_0 (mn); N_n : Highest Biomass Concentration $(mg.L^{-1})$ corresponding to time t_n (mn).

II.1. Physico-chemical analyses

II.1.1. Determination of dry matter

To determine dry matter (DM) content of wet algal "pastes", a standard MB35 moisture meter was used. This equipment works on the principle of thermogravimetry, which consists in measuring the mass variation of a sample according to the temperature, which is fixed to 110°C until a constant mass was obtained [17]. Once the stabilized weight has been reached, drying is completed and the result displayed indicates the percentage of moisture present in the matrix and the final weight of the sample. Dry matter calculation of the algal paste was carried out as follows:

$$DM\% = 100 - HM\%$$

Avec : DM%: percentage of dry matter ; HM%: pourcentage d'humidité donné par l'humidimètre

II.1.2. Lipid extraction

Lipid extraction from dry Oedogonium nodulosum algae was performed using the Soxhlet system (ISO 659-1988) [18]. This method consists in extracting oils contained in the algal cells below 70°C using for our case a combination of two organic solvents: n-hexane and chloroform (2: 1). The solvent, contained in the flask, is boiled, which then transfers it to the upper part of the Soxhlet (Fig. 2). There, it is condensed by a refrigerant at the top of the installation and accumulates around and inside the cartridge. When the solvent reaches the upper level of the siphon, the mixture is returned to the tank by pressure difference, where it is evaporated again. Several extraction cycles are thus carried out, the duration of the operation is fixed at 4 hours according to the standard [18]. It is then considered to have reached total solute depletion of the solid substrate and concentrated the extract.



Fig. 2 Soxhlet diagram (source: Engineering Tech.) R : Refrigerant, E : Extractor, B : Balloon

The extracted oil was then placed in the Rotavapor flask to remove organic solvents in excess (Fig. 3) by distillation process at 70°C with 100 rpm [19].



Fig. 3 Rotavapor type R3000-Gemini BV for the distillation of extracted oil

II.2.1. Homogeneous transesterification reaction

A basic homogeneous transesterification reaction was conducted to transform the extracted oil into an ester [20]. 200 ml intake of oil extracted from Oedogonium nodulosum is placed in a tricols flask, to which 341 ml of methanol (MeOH) and 1.85 g of KOH were added. The proportion of the catalyst amount to be used varies within a range of 0.5 to 1% of the oil mass, for a conversion efficiency of 94 to 99% [14, 20]. The mixture is heated up rapidly to 70°C, for the reaction under mechanical agitation of 100 rpm [21]. The standard transesterification reaction is as follows:

$Triglyceride + Alcohol \leftrightarrow Ester alkylique + Glycerol$

However, the molar ratio alcohol/oil 9/1 chosen for our case of methanolic transesterification of the extracted oil, favors a better separation of glycerin because it increases its

solubility [22]. Methanol is the alcohol used in our transesterification reaction, due to its better reactivity than ethanol [19, 22]. This efficiency advantage is that the use of methanol does not pose any difficulties during separation [20]. The water content of the extracted oil is 0.5 (% by weight), and the total phosphorus content was 0.8 ppm.

After 120 minutes of stirring, the resulting mixture is poured into a separating funnel and left to stand for 24 hours, in order to promote the separation of the biofuel from Glycerol/methanol.

The transition from an oil state to an alkyl ester state, commonly known as biodiesel, makes it possible to: reduce the molecular weight to one-third of that of the oil, reduce the viscosity by a factor of 8, reduce the density and increase its volatility [24, 25].

A purification step of the ester consists in removing impurities such as residual glycerin, excess alcohol, traces of catalysts, soaps and salts formed by the homogeneous catalysis. This purification operation is carried out by flash distillation under vacuum.

After washing the biodiesel with water, the drying operation allows the water present to be removed by heating the product to a high drying temperature $(120^{\circ}C)$ in the oven.

III. RESULTS & DISCUSSION

A. Biomass Growth

Based on microscopic examination, we mainly observed the abundance of chlorophyceae, and the culture conditions in the photobioreactor in batch mode showed a concentration of algal biomass of the *Oedogonium nodulosum* species, with the higher growth rate ($\mu_{max} = 4.9 \text{ days}^{-1}$) (Fig. 4).





From the maximum growth rate obtained, we can deduce that the maximum number N of Oedogonium nodulosum was reached from the 11th day of pure algal culture.

$$Log_2N_n = X = 41.5$$
 where : $N = 2^X = 2^{41.5} \approx 10^{18} \text{ cells.L}^{-1}$.

B. Characterization of the Extracted Oil

This cultured algae paste was used for lipid extraction. After drying, we obtained a dry mass of *Oedogonium nodulosum* equal to 259.87g from a wet paste of 1.55kg. The mass of the extracted oil from this dry algal mass is 112.58g with an extraction efficiency rate of 43.32%.

C. Transesterification Reaction Efficiency

The influence of various transesterification reaction times (1, 2 hours) at temperature (70°C) was studied (mass ratio MeOH / oil of 9 /1 and 1% by mass KOH). The evolution of biodiesel yield (g biodiesel /g oil) over the time of reaction is shown in (Fig. 5).



Fig. 5 Evolution of biodiesel yield according to reaction time (mass ratio MeOH/oil: 9/1 KOH : 1% mass, 70° C)

The transesterification reaction results show that the maximum biodiesel yield of 80% is obtained after 2 h reaction is complete. An efficiency of 70% is obtained after 1 hour of reaction at room temperature. Therefore, the reaction time seems to have an influence on biodiesel yield.

D. Physicochemical Properties of Biodiesel

The physico-chemical properties of biodiesel produced from *Oedogonium nodulosum* oil, compared to those of diesel fuel, are presented in Tab. 1.

Tab. 1 Physicochemical properties of biodiesel from *Oedogonium nodulosum* oil compared to diesel oil

Compounds	рH	Density g.cm ⁻³	Viscosity at 40°C mm ² .s ⁻¹	Flash point °C	Cloud point °C
Oedogonium nodulosum oil	8.59	0,9236	0,663	155	-10
Biofuel	9.77	0,8484	1,8984	138	-12
Diesel Fuel	9.39	0,8393	2,8620	8	-14

Biodiesel has a lower density, viscosity and flash point than extracted oil. These lower physico-chemical properties of biodiesel compared to that of the oil correspond to those of methyl esters by glycerol removal. However, the density and flash point of biodiesel are higher than those of diesel. These results show that the combustion of the latter, particularly at start-up, is easier than that of biodiesel.

E. Optimization of Biodiesel Yield

In order to determine the optimal biodiesel yield, a series of transeterification reactions of the extracted oil were carried out under the following conditions: reaction time: 1h, temperature 70°C, mass ratio Methylene / oil: 12/1, 9/1, 6/1 with a mass percentage KOH / oil: 1%.

Tab. 2 Biodiesel production results according to the alcohol/oil mass ratio

	1st trial	2nd trial	3rd trial
Alcohol / oil mass ratio	12/1	9/1	6/1
Yield (%)	79	80	68

The best separating result was obtained with a ratio of 9/1 (Fig. 6).



Fig. 6 Biodiesel produced by transesterification separated from the organic phase Glycerol/methanol

Glycerol is essentially insoluble in biodiesel, so almost all glycerol is easily removed by settling. Alcohol can act as a cosolvent to increase the solubility of glycerol in biodisel. In our case, the yield obtained is less than 100 (Tab. 2). This can be explained by the fact that glycerol has not been solubilized in biodiesel due to the transformation of glycerol into fatty acid which in turn has undergone saponification in the presence of KOH [26, 27, 28]. Indeed, during transesterification, parallel parasitic reactions occur at the same time [28, 29, 30]. Fatty acids react with a basic catalyst to form soap [31, 32]. This will inevitably reduce the conversion efficiency [33].

IV. CONCLUSION

From the standpoint of renewable energy and sustainable biofuels development, third-generation are attracting increasing investment interest from researchers and some green industries. Biofuels are emerging as an essential transport energy source. One of the interesting sources of biofuels are algae, which are capable of producing biodiesel and bioethanol that can be used in different types of engines. Biodiesel, which is derived from triglycerides methyl or ethyl alcohol by transesterification, has attracted considerable attention over the past three decades due to its benefits: renewable, biodegradable and non-toxic fuel. The objective of this work was to synthesize a biodiesel from green filamentous microalgae "Oedogoium nodulosum" oils, by transesterification, which is the most widely used technique for biodiesel synthesization. For this purpose, we used a shortchain alcohol (methanol) for the reaction and KOH as a catalyst. Parameters affecting alkyl ester yield such as catalyst mass percentage (1% KOH) and catalyst/oil mass ratio were studied over time.

The highest biodiesel yield obtained is 80% and corresponds to a mass ratio of 9/1 Methanol / oil, at a temperature of 70° C and a reaction time of 2 hours.

V. REFERENCES

- [1] A Préat (2012). Paniquer sur les réserves du pétrole ? Université libre de Bruxelles. Available : http://www.academieroyale.be/academie/documents/AlainPreatPaniquere servespetrole15844.pdf
- [2] Keniar, K., Ghali, K., & Ghaddar, N. (2015). Study of solar regenerated membrane desiccant system to control humidity and decrease energy consumption in office spaces. *Applied Energy*, Volume: 138:121-132.
- [3] D. Kramer (2017). Biology lead the race to turn sunlight into fuels. *Physics today*, 70, 4, 30.
- [4] A. Demirbas and M. Demirbas (2009). Algae Energy: Algae as a New Source of Biodiesel. *Springer*, 224.
- [5] B. S. SAHARAN, D. SHARMA, R. SAHU, O. SAHIN and A. WARREN (2013). TOWARDS ALGAL BIOFUEL PRODUCTION: A CONCEPT OF GREEN. *Innovative Romanian Food Biotechnology*, 12: 1-21.
- [6] B. SANDESH KAMATH (2007). Biotechnological production of microalgal carotenoids with refrece to astaxanthin and evaluation of its biological activity. *Thèse de doctorat, Univ of Mysore, India*
- [7] R. Voloshin, M. V. Rodionova, S. K. Zharmukhamedov, S. I. Allakhverdiev (2016). Review: Biofuel production from plant and algal biomass. *International Journal of Hydrogen Energy* 41(39). DOI: 10.1016/j.ijhydene.2016.07.084
- [8] W. RYAN and A. Z. HUNT (2009). Das Electromagnetic Biostimulation of Living Cultures for Biotechnology, Biofuel and Bioenergy Applications 10, 4515-4558. *Int. J. Mol. Sci.*, 10: 4515-4558.
- [9] OFID study prepared by IIASA (2009). Biofuels and food security. Implications of an accelerated biofuels production. Vienna, Austria. Available: <u>http://www.globalbioenergy.org/uploads/media/0903_OFID_BiofuelAndFoodSecurity.pdf</u>
- [10] J. T. Teck Chye, Y. J. Lau, L. S. Yon, M. K. Danquah, (2018). Biofuel production from algal biomass. In book: Bioenergy and Biofuels Edition: First Chapter: 3 Publisher: CRC Press, Boca Raton (Taylor & Francis Group) Editors: Ozcan Konur. DOI: 10.1201/9781351228138-3
- [11] Y. Chisti (2007). Biodiesel from microalgae. *Biotechnology Advances*, 25: 294–306.
- [12] E. I. Bello, T. S. Mogaji and M. Agge (2011). The effects of transesterification on selected fuel properties of three vegetable oils. *Journal of Mechanical Engineering Research*, 3(7): 218-225.
- [13] S. Nasreen, M. Nafees, L. A. Qureshi, M. Shahbaz Asad, A. Sadiq and A. S. Danial (2018). Review of Catalytic Transesterification Methods for Biodiesel Production. *Open access peer-reviewed chapter*, DOI: 10.5772/intechopen.75534.
- [14] A. P. Vyas, P. H. Shukla and N. Subrahmanyam (2011). Production of Biodiesel using Homogeneous Alkali Catalyst and its Effect on Vehicular Emission. *INTERNATIONAL CONFERENCE ON CURRENT TRENDS IN TECHNOLOGY*, 1-7.
- [15] J.C. DRUART et F. RIMET. Protocoles d'analyse du phytoplancton de l'INRA : prélèvement, dénombrement et biovolumes. INRA-Thonon, Rapport SHL 283 - 2008, 96 p.
- [16] A. E. Simosa (2016). Factors affecting algal biomass growth and cell wall destruction. University of New Orleans Theses and Dissertations. 2277. Available: <u>https://scholarworks.uno.edu/td/2277</u>

[17] C.J. Zhu and Y.K. Lee (1997). Determination of biomass dry weight of

marine microalgae Journal of Applied Phycology (1997) 9: 189. Available:

- https://doi.org/10.1023/A:1007914806640
- [18] ISO 659: 1988 (1988). Oilseeds -- Determination of hexane extract (or light petroleum extract), called "oil content". International Organization for Standardization. Available: <u>https://www.iso.org/standard/4815.html</u>
- [19] C. Dejoye (2013). Eco-extraction et analyse de lipides de micro-algues pour la production d'algocarburant. Université d'Avignon, NNT : 2013AVIG025.

- [20] M. K. Lam, K. T. Lee, and A. R. Mohamed (2010). Homogeneous, heterogeneous and enzymatic catalysis for transesterification of high free fatty acid oil (waste cooking oil) to biodiesel: A review. *Biotechnology Advances*, 28: 500-518.
- [21] S. PAZ, P. Meneghetti, MR Meneghetti, CR. Wolf (2007). Transformation of triglycerides into fuels, polymers and chemicals: some applications of catalysis in oleochemistry. Quim Nova 2007; 30:667–76.
- [22] Y. Tang, Y. Zhang, J. Rosenberg, F. Wang (2016). Optimization of One-Step *In Situ* Transesterification Method for Accurate Quantification of EPA in *Nannochloropsis gaditana*. *Applied Sciences* 6(11):343.
- [23] G. R. Stansell, V.M. Gray and S.D. Sym (2012). Microalgal fatty acid composition: implications for biodiesel quality. *Journal of Applied Phycology*, volume 24, numéro 4, p.791-801.
- [24] A. GERAR TCHAKBLO (2009). Production de Biodiesel par Tansestérification Alcoolique : étude sur réacteur pilote, pp 24.
- [25] M. Hunsom, P. Saila, P. Chaiyakam and W. Kositnan (2013). Comparison and Combination of Solvent Extraction and Adsorption for Crude Glycerol Enrichment. *INTERNATIONAL JOURNAL of RENEWABLE ENERGY RESEARCH*, Vol.3, No.2, 2013.
- [26] B. K. Uprety (2017). Conversion of crude glycerol from the biodiesel industry to value added products. Lakehead University, Thunder Bay, Ontario, Canada, 215 p.
- [27] J. A. Dutton (2015). Alternative Fuels from Biomass Sources. e-Education Institute. <u>https://www.e-education.psu.edu/egee439/node/684</u>
- [28] T. Rachmanto, D. Allanson, C. Matthews and I. Jenkinson (2014). Monitoring of Biodiesel Transesterification Process Using Impedance Measurement. International Journal of Materials, Mechanics and Manufacturing, Vol. 2, No. 4.
- [29] A. Rodrigues, J. C. Bordado and G. dos Santos, (2017). Upgrading the Glycerol from Biodiesel Production as a Source of Energy Carriers and Chemicals— A Technological Review for Three Chemical Pathways. Energies, 10, 1817.
- [30] F. Trejo-Zárraga, F. Herernádez-Loyo, J. C. Chavarría-Hernández and R. Sotelo-Bovás (2017). Kinetics of Transesterification Processes for Biodiesel Production. Intech Open Chapter Book. DOI: 105772/intechopen.75927.

[31] E. Suyenty, H. Sentosa, M. Agustine, S. Anwar, A. Lie and E. Sutanto,

(2007). Catalyst in Basic Oleochemicals. Bulletin of Chemical Reaction

Engineering & Catalysis, 2(2-3), 22-31.

- [32] K. Amri, I. Paryanto, A. Kismanto, Adiarso and T. H. Soerawidjaya (2011). Esterification Reaction of High Free Fatty Acid Content of Crude Palm Oil and Glycerol Using Zinc Glycerolate and Zinc Soap Catalysts. Conference: Sriwijaya International Seminar on Energi Science and Technology. DOI: 10.13140/RG.2.1.2431.4409
- [33] G. Yoo, M. S. Park and J. W. Yang, (2015). Chemical Pretreatment of Algal Biomass. *In book* Pretreatment of Biomass. DOI: 10.1016/B978-0-12-800080-9.00012-8.